consulting researchers based there. When leaving a country, flies in my checked baggage have never received any interest at the airport. Once in Ethiopia, a baggage scanner wanted to look at the empty tubes of fly food in my wife's bag because she thought it looked like a carpet (perhaps those are subject to export tax), and was then suspicious about the tubes, but eventually they let us through (after I tasted the instant fly food in front of them and assured them that it was "flakes" and not "powder"). When shipping flies, convincing an African DHL office to accept a shipment of live flies is not guaranteed, but my success rate has been >90%. The primary documents of interest to DHL have been my U.S. import permit and an official-looking letter from my home institution). For bringing or shipping flies to the U.S., an import permit from the USDA PPQ office is needed, and can be applied for online. Note that if you're flying in with flies: you should let the USDA know when you're coming, you have to enter the U.S. at the "port of entry" listed on your permit, you must have one of your permit's "mailing labels" with the flies, and the flies should be sealed inside a second box.

Armchair collecting:

In case you'd like to have some flies collected but no one is available to make the trip, I've also had fairly good luck contacting researchers in Africa and asking if they'd be willing to send me some flies. Basically, I sent these kind people some simple collection equipment, compensated their travel expenses (a few hundred dollars at most), and they shipped flies to me. Results were variable but generally good, especially when I could find an entomologist to help. The instructions I sent to them resemble the methods described above. An alternative to the isofemale line preparation for less experienced collectors (which I have not yet tested) would be to receive a mixed-sex batch of flies, age and transfer the wild-caught females until they run out of sperm, and then pair each with a different wild-caught male.

Correction

Johnson, David A. 2007. Amplification of DNA from 30-year-old aceto-orecein stained salivary gland squash slides. Dros. Inf. Serv. 90: 156-158.

Two corrections are highlighted in bold below in the procedure steps 23 and 26. Step 23 should indicate –70°C, not 70°C; and step 26 omitted 70% before the word ethanol.

23) Incubate at **–70°**C for 5 minutes

26) Add 1000 µl **70%** ethanol, mix gently, and centrifuge for 1 minute at 13,000-16,000 ×g.

Call for Papers

Submissions to Drosophila Information Service are welcome at any time. The annual issue now contains articles submitted during the calendar year of issue. Typically, we would like to have submissions by mid-December to insure their inclusion in the regular annual issue. but articles can be accepted for this volume until 31 December. Details are given in the Guide to Authors or on the DIS web site: www.ou.edu/journals/dis.